RESEARCH

Open Access



Shifted phase of EEG cross-frequency coupling in individuals with Phelan-McDermid syndrome

Michael. G. Mariscal¹, Elizabeth Berry-Kravis^{2,3,4}, Joseph D. Buxbaum^{5,6,7,8}, Lauren E. Ethridge⁹, Rajna Filip-Dhima¹, Jennifer H. Foss-Feig⁵, Alexander Kolevzon^{5,6}, Meera. E. Modi¹, Matthew W. Mosconi¹⁰, Charles A. Nelson¹¹, Craig M. Powell¹², Paige M. Siper⁵, Latha Soorya¹³, Andrew Thaliath², Audrey Thurm¹⁴, Bo Zhang¹, Mustafa Sahin¹ and April R. Levin^{1*} on behalf of the Developmental Synaptopathies Consortium

Abstract

Background: Phelan-McDermid Syndrome (PMS) is a rare condition caused by deletion or mutation of the *SHANK3* gene. Individuals with PMS frequently present with intellectual disability, autism spectrum disorder, and other neurodevelopmental challenges. Electroencephalography (EEG) can provide a window into network-level function in PMS.

Methods: Here, we analyze EEG data collected across multiple sites in individuals with PMS (n = 26) and typically developing individuals (n = 15). We quantify oscillatory power, alpha-gamma phase-amplitude coupling strength, and phase bias, a measure of the phase of cross frequency coupling thought to reflect the balance of feedforward (bottom-up) and feedback (top-down) activity.

Results: We find individuals with PMS display increased alpha-gamma phase bias (U = 3.841, p < 0.0005), predominantly over posterior electrodes. Most individuals with PMS demonstrate positive overall phase bias while most typically developing individuals demonstrate negative overall phase bias. Among individuals with PMS, strength of alpha-gamma phase-amplitude coupling was associated with Sameness, Ritualistic, and Compulsive behaviors as measured by the Repetitive Behavior Scales-Revised (Beta = 0.545, p = 0.011).

Conclusions: Increased phase bias suggests potential circuit-level mechanisms underlying phenotype in PMS, offering opportunities for back-translation of findings into animal models and targeting in clinical trials.

Keywords: Phelan-McDermid syndrome, EEG, Power, Cross-frequency coupling, Phase bias

Introduction

Phelan-McDermid Syndrome (PMS) is a well-characterized genetic condition that results from haploinsufficiency of *SHANK3* in the 22q13.3 region. The phenotype in PMS is frequently characterized by intellectual disability [1, 2], autism spectrum disorder (ASD; 50–84%) [3, 4],

*Correspondence: April.Levin@childrens.harvard.edu

¹ Department of Neurology, Boston Children's Hospital, Boston, MA, USA

Full list of author information is available at the end of the article

and epilepsy [5]. *SHANK3* codes for a master scaffolding protein in the postsynaptic density of glutamatergic synapses [6], and its isoforms perform a variety of synaptic functions relevant to neuronal excitability and plasticity [7-15].

A key step in understanding the translational pathway from cells to circuits, networks and ultimately phenotype, involves measurements that reflect large scale network dynamics, including assessments of intrinsic neural oscillations. Electroencephalography (EEG) offers



© The Author(s) 2021. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/. The Creative Commons Public Domain Dedication waiver (http://creativecommons.org/publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

particular opportunity in this regard, because it can measure network dynamics in both humans and animal models, allowing for both forward and back-translation of findings. Clinical EEG (evaluated by visual review) frequently demonstrates abnormalities in PMS, including generalized slowing of activity, slowing or absence of the occipital dominant rhythm, and epileptiform activity [5, 16]. Epileptiform activity on EEG is also frequently seen in ASD more broadly [17]. Shank3B null mutant mouse models have demonstrated altered oscillatory power, depending on the location and frequency band studied [9, 14, 18]. Numerous studies in humans with ASD have demonstrated various abnormalities in resting EEG spectral power [19]; however, quantitative studies of EEG activity in humans with PMS have not been previously published.

Recently, there has been increasing interest in the coupling of EEG activity across frequencies, using measures such as Phase-Amplitude Coupling (PAC), given the possibility that such cross-frequency coupling has distinct mechanistic underpinnings. Such coupling is crucial for many of the cognitive functions that are altered in neurodevelopmental disorders, such as long-range communication [20], integration of local and global cortical processing [21], and segmenting and prioritizing sensory input [20, 22]. Altered PAC strength in the alpha-gamma frequency pair has been reported in individuals with ASD at baseline (i.e., rest) and associates with symptom severity [23, 24]. PAC is also altered during tasks in some neurodevelopmental disorders, including during face processing in ASD [25] and cognitive discrimination in a mouse model of Fragile X syndrome [26]. Likewise, PAC during the period preceding an auditory stimulus has been found to positively correlate with non-verbal intelligence quotient in Fragile X syndrome [27]. Cross-frequency coupling thus has theoretical relevance to ASD, intellectual disability, and associated neurogenetic disorders in which such processes are likely altered [28–30].

Recent work suggests not just the strength (the extent to which PAC occurs), but the phase (e.g., where in relation to the alpha waveform gamma amplitude is maximal), can signal important network characteristics. The phase at which fast oscillations are strongest can vary by cortical layer [31] and with alterations in interneuron function [32]. Surface EEG measurements demonstrate the alpha phase resulting in maximum gamma power can vary by age [33] and depth of anesthesia [34]. Differences in PAC phase have been found with encoding success [35] and context [36], and PAC phase bias has been suggested to reflect the ratio of feedforward (bottom-up) to feedback (top-down) cortical activity [33], suggesting the phase of PAC can be functionally relevant particularly among conditions commonly associated with autism spectrum disorder. The timing of gamma within the alpha cycle consequently has the potential to capture alterations in brain connectivity and function that result from specific synaptic perturbations and underlie clinical disorders.

EEG measures of PAC strength and PAC phase thus offer opportunities to enhance understanding of circuit-level dysfunctions in PMS. Here, we first examined whether individuals with PMS, as compared to typically developing (TD) individuals display differences in alphagamma PAC strength and phase. Second, we investigated whether these EEG metrics associate with measures of phenotype among individuals with PMS. We hypothesized (1) individuals with PMS would demonstrate increased PAC strength and phase bias, compared to typically developing controls and (2) PAC metrics would correlate with sensory processing difficulties and ASD symptom severity.

Methods

Participants

Participants were recruited through a prospective, observational cohort study at four institutions across the United States as a part of the Developmental Synaptopathies Consortium (Clinical Trial NCT02461420): Icahn School of Medicine at Mount Sinai, University of Texas Southwestern, Rush University Medical Center, and Boston Children's Hospital. Stanford University and the National Institute of Mental Health also participated in the overarching study, but because they only collected phenotyping data and did not collect EEG, participants recruited at those institutions are not included here. In total, 31 individuals with PMS and 17 TD individuals had EEG completed. Participants with PMS were included if they had pathogenic deletions or mutations of the SHANK3 gene; clinical reports were reviewed to confirm this information. Typically-developing individuals were matched at the group level with PMS participants on chronological age and sex. TD individuals were excluded if they had a diagnosis of any intellectual disability, ASD, or other learning, developmental, psychiatric, or neurological disorders as determined by parent report. All participants were 4 to 19 years of age (inclusive). Informed written consent was obtained from legal guardians and assent was obtained from participants when appropriate. Table 1 shows demographics for participants with adequate EEG data for inclusion (see below).

Phenotypic data

To examine how our EEG measures related to developmental abilities and ASD phenotypes among individuals with PMS, the following assessments were conducted: the Vineland Adaptive Behavior Scales (Vineland II):

Table 1 Demographic information for participants with useableEEGs

	PMS (n = 26)	TD (n = 15)
Sex	10 (M) 16 (F)	9 (M) 6 (F)
Collection site*		
Icahn School of Medicine at Mount Sinai	11 (42.3%)	10 (33%)
Rush University Medical Center	8 (30.8%)	5 (67%)
Boston Children's Hospital	7 (26.9%)	0 (0%)
Net type		
Hydrocel 128	18 (69%)	10 (33%)
Biosemi 32	8 (31%)	5 (67%)
Age (years)	9.5 ± 4.25	10.0 ± 2.39
ASD diagnosis		
ASD	11 (42%)	0 (0%)
Non-ASD	14 (54%)	15 (100%)
Unknown	1 (4%)	0 (0%)
ADOS		
Completed	22 (85%)	0 (0%)
Comparison score	6.05 (2.54)	-
Vineland		
Completed	25 (96%)	0 (0%)
Adaptive behavior composite standard score	52.8 ± 13.4	-
Communication composite standard score	58.0 ± 14.6	-
SSP		
Completed	22 (85%)	3 (20%)
Total score	143.9 ± 16.5	178.7 ± 17.1
RBS-R		
Completed	23 (88%)	0 (0%)
Total score	16.26 ± 15.7	-
MSEL		
Completed	16 (61.5%)	0 (0%)
NVIQ	19.98 ± 10.9	-
SB5		
Completed	9 (34.6%)	0 (0%)
NVIQ	46.56 ± 7.5	-
DAS		
Completed	2 (7.7%)	0 (0%)
NVIQ	$73.50 \pm .71$	-
NVIQ		
Completed	25 (96%)	0 (0%)
Non-verbal intelligence quotient	31.4 ± 17.9	-
Seizure history		
Yes	4 (15.4%)	-
No	21 (80.8%)	-
Unknown	1 (3.8%)	15 (100%)
SHANK3	-	
Mutation	7 (26.9%)	-
Deletion	19 (73.1%)	-
Deletion size (mega base pairs)	3.95 ± 3.0	-

Categorical variables (i.e. yes, no) are presented as the number in each category, followed by the percentage in each category. Continuous variables are presented as the mean value \pm their standard deviation. *No EEGs from University of Texas Southwestern retained enough data after artifact rejection

Table 1 (continued)

to be analyzed

Survey Interview Form [37], the Autism Diagnostic Observation Schedule, 2nd edition (ADOS-2) [38], the Autism Diagnostic Interview-Revised [39], the Autism Diagnostic Criteria Checklist from the Diagnostic and Statistical Manual of Mental Disorders, 5th edition [40], the Short Sensory Profile (SSP) [41], and the Repetitive Behavior Scale-Revised (RBS-R) [42]. A psychologist determined ASD diagnosis either on the basis of the study's assessments or clinical experience when the participant was seen clinically on a regular basis. Additionally, to assess non-verbal cognitive ability, participants were either given the Mullen Scales of Early Learning (MSEL) [43] the Stanford Binet-5 (SB-5) [44], or the Differential Ability Scales, 2nd edition (DAS-II) [45]. Data for a non-verbal intelligence quotient (NVIQ) was compiled depending on the test given: for participants given the SB-5, NVIQ was taken; for participants given the MSEL, the mean of the visual reception developmental quotient (visual reception age equivalent score/age in months) and fine motor developmental quotient (fine motor age equivalent score/age in months) was taken; for participants given the DAS-II, the non-verbal reasoning standard score was taken. Finally, for participants who had experienced seizures, a seizure history was collected.

EEG acquisition/processing

Continuous EEG was collected for up to 10 min. Participants viewed a silent movie of their choice during EEG recording as is common practice in individuals with neurodevelopmental disorders [47]. EEG was recorded using either a 128-channel Hydrocel Geodesic Sensor Net or a 32 channel ActiveTwo Biosemi net. Data were sampled at either 512 Hz or 1000 Hz (all files were later resampled to 250 Hz). Impedances were kept below the recommendations for the specific EEG system being used prior to recording. For a subset of individuals with PMS, continuous EEG was again collected approximately 12 weeks after the initial recording. These subsequent recordings were used in place of initial recordings if the initial recording did not meet data quality thresholds (n=1); all other analyses were performed using the initial recording.

Files were processed using the Batch EEG Automated Processing Platform (BEAPP) [48]. Within BEAPP, the Harvard Automated Preprocessing Pipeline for EEG (HAPPE), which was developed specifically to optimize preprocessing of developmental EEG data with potentially high levels of artifact and short recordings, was used to automate preprocessing and artifact minimization

[49]. Data were first filtered using a 1 Hz high-pass filter and a 100 Hz low-pass filter. Data were then downsampled to 250 Hz for optimal performance of the HAPPE pipeline. With the exception of Cz, which was used as a reference electrode in some sites' systems, only electrodes in the international 10-20 system were included in this analysis (18 total) to allow standardization of analyses across net types. Epochs of signal with any channel's amplitude>40 µV (the HAPPE default threshold, reflecting the reduced signal amplitude that results from wavelet-thresholding and independent components analysis in HAPPE) were removed. EEG recordings were removed from further analysis if they exceeded thresholds for HAPPE data quality as per [50] in one or more of the following output parameters: percent good channels, mean retained artifact probability, median retained artifact probability, percent of independent components rejected, and percent variance retained after artifact removal. Data were subsequently re-referenced using an average reference, and then segmented into 2 s windows for power and PAC analysis. For each participant, 150 segments (300 s of data) were randomly selected; files with fewer than 150 segments of data at this stage were not analyzed. Primary power and PAC metrics were then obtained using code added to the BEAPP software.

Power analyses

Power was computed across frequencies using a three taper multitaper window [51]. Power was then computed for a number of frequency bands: Delta [1–4 Hz), Theta [4–8 Hz), Alpha [8–12 Hz), Beta [12–30 Hz), and Gamma [30–55 Hz). Total power was computed as all frequencies between [1–55 Hz]. The power at each frequency band and the overall 1–55 Hz range was computed by adding the power spectral density over the frequency range of interest.

To capture each frequency band's relative contribution to total power, the relative power at each frequency band was computed as the power at each frequency band divided by the total power. Power values were then averaged across electrodes. Visual inspection of the power spectra, averaged across the occipital channels analyzed in this study (O1 and O2), was used to identify the peak alpha frequency of each participant.

PAC analysis

Modulation index

To capture the presence of coupling, PAC was first quantified using the Modulation Index (MI) [52]. Because the data are not time locked to any specific task, we focus on PAC in the alpha-gamma range, where prior studies have shown abnormalities in other neurodevelopmental disorders using resting or non-time-locked data [23, 24]. For each frequency pair, the raw signal in each segment was exported from MATLAB into Python and filtered into a range of alpha (8-12 Hz in 2 Hz steps) and gamma (here, 28-56 Hz to allow for division into 4 Hz steps) frequencies using code adapted from Dupré la Tour et al. [53]. Alpha frequencies were filtered using a constant bandwidth of 2 Hz, while gamma frequencies were filtered using an upper sideband variable bandwidth, so as to avoid including phase frequencies in the amplitude frequencies. In detail, for each gamma frequency, the lower passband cutoff was 2 Hz below the gamma frequency, and the upper passband cutoff was set as the alpha phase frequency plus the gamma amplitude frequency [24]. For example, for the combination of 40 Hz gamma and 8 Hz alpha, the lower limit of the gamma amplitude filter was 38 Hz (40-2), while the upper limit of the filter was 48 Hz (40+8). Filtering at this step consisted of a zero-phase cosine-based filter to extract the real component, and then a sine-based filter to extract the imaginary component, resulting in a complex-valued output signal [53]. The alpha phase time series, or gamma amplitude time series, were obtained from this complex signal. The phases of the alpha signal were then binned into 18 20° intervals (-180° to 180°), and the mean of the amplitude of the gamma signal occurring within each phase bin was calculated. Mean gamma amplitude values in each phase bin were then normalized by dividing each bin value by the sum of all bin values. Data were then imported into MATLAB, where the amplitude of the gamma signal at each phase bin of the alpha signal was then averaged together across segments. The MI_{raw} was then computed as the Kullback-Leibler divergence of the gamma amplitude distribution from a uniform distribution [52]. We then employed a time-shift procedure to control for factors that may generate spurious phaseamplitude coupling. In detail, for each participant, 200 surrogate MI values (MI_{surr}) were generated by repeating the procedure after offsetting gamma amplitude from the alpha phase distribution by a randomized time shift between 0.1 and 1.9 s. A normalized MI (z-MI) was then computed as the z-score of the $\ensuremath{\text{MI}}_{\ensuremath{\text{raw}}}$ compared to the distribution of MI_{surr} values [54]. The z-MI at each alpha and gamma frequency combination was then averaged to obtain a single overall alpha-gamma PAC value for each participant, at each electrode.

Phase bias

Modulation Index captures the extent of coupling. We additionally set out to quantify whether gamma amplitude increased closer to the rising or falling phase of the alpha waveform, and to what degree. To do so, we employed a metric termed phase bias, drawing on measures of phase preference [32] and prior findings that this

tends to show a bimodal distribution (i.e., with fast activity occurring maximally at either the negative or positive phases (corresponding to rising and falling phases, respectively) of the slower waveform [35]. Specifically, we quantified the phase bias of the gamma amplitude to the positive phases of the alpha waveform; i.e., the relative change of gamma amplitude (gamma_{amp}) during the positive phases (0°-180°) of the alpha waveform. Thus, phase bias is calculated as (Σ gamma_{amp} in positive phases of the alpha waveform)/ $\Sigma(\text{gamma}_{\text{amp}}$ in all phases of the alpha waveform) -0.5. Importantly, here, a cosine-based filter was used to extract alpha phase; as a result, 0° corresponds to the peak of $alpha, +90^{\circ}$ corresponds to the falling zero crossing of alpha, 180°/-180° corresponds to the trough of alpha, and -90° corresponds to the rising zero crossing of alpha. Therefore, a phase bias>0 indicates gamma amplitude increases at the falling phase of the alpha waveform, and a phase bias < 0 indicates gamma amplitude increases at the rising phase of the alpha waveform. Additionally, a larger distance from 0 (where gamma amplitude does not increase preferentially at either positive or negative phases of alpha) indicates stronger phase bias. The phase bias at each alpha frequency and gamma high frequency combination was then averaged to obtain a single overall alpha-gamma phase bias value.

Statistical analysis Group comparisons

We first set out to test whether power or phase-amplitude coupling metrics differed between groups. Because most metrics were not normally distributed, all group comparisons were performed using a non-parametric test (independent samples Mann-Whitney U) unless otherwise specified. Relative power in each frequency band was compared between groups, and an independent samples t test was used to compare peak alpha frequency between groups. To test whether overall PAC metrics differed in individuals with PMS as compared to typically developing individuals, group comparisons were first performed on z-MI and phase bias data averaged across all 10-20 electrodes. Subsequently, because PAC has been shown to differ between anterior and posterior scalp areas [33], these group comparisons were repeated after averaging PAC metrics across all anterior 10-20 electrodes (Fp1, Fp2, F3, F4, F7, F8, Fz) and then posterior 10-20 electrodes (P3, P4, P7, P8, Pz, O1, O2). Finally, these comparisons of overall, anterior, and posterior z-MI and phase bias were repeated between individuals with PMS diagnosed with ASD (N = 11), and individuals with PMS diagnosed without ASD (N = 14). Data were analyzed in SPSS (IBM Corp, 2016).

Clinical associations

All associations were performed using linear regression analysis. Because PAC has been shown to change with age [33], we tested whether age was associated with PAC metrics among all participants. Additionally, to test whether the relationship between ln(z-MI) and age was different in individuals with PMS as compared to TD individuals, a regression was performed, with ln(z-MI) as the dependent variable, and age, group, and age by group included as independent variables. The association between alpha power and PAC metrics (averaged across all electrodes) was additionally examined. To test how PAC associated with behavioral phenotype in individuals with PMS, linear regression analysis was performed between PAC metrics (z-MI and phase bias) and the following measures: Vineland Adaptive Behavior Composite, Vineland Socialization Composite, ADOS comparison score, SSP, RBS-R, and NVIQ. Additional linear regressions were performed between z-MI and the 6 behavior sub-scales of the RBS-R (Restricted Interest, Sameness, Ritualistic, Compulsive, Self-Injurious, and Stereotypic). Because z-MI did not demonstrate a normal distribution, linear regressions were performed on the natural log transformation of z-MI; one negative z-MI value was not included in this analysis. Age was included as a control variable in all regressions.

Clinical comparisons

In individuals with PMS, we tested whether PAC measures differed by a number of categorical clinical variables, including: sex, presence of an ASD diagnosis, presence of a seizure history (at least one seizure event experienced), and whether the participant has a *SHANK3* mutation or deletion. All comparisons were computed using a Mann– Whitney U test. For all associations between EEG and clinical measures, a Benjamini–Hochberg correction was applied to power, ln(z-MI) and phase bias clinical correlations separately (FDR=0.1).

Participants with insufficient EEG data

In total, there were 33 individuals with PMS and 17 TD individuals in the study. After removal of participants who did not complete EEG (n = 2 with PMS), had insufficient data quality (n = 4 with PMS, n = 1 TD), or had insufficient data length (n = 1 with PMS and n = 1 with TD), 26 individuals with PMS and 15 TD individuals remained for further analysis. Compared to PMS participants included in this dataset, the 7 PMS participants excluded for unusable EEG data were more likely to be male (6/7). Otherwise, they were not significantly different in age (mean = 9.94, SD = 4.58, p = 0.7587), they demonstrated similar prevalence of ASD diagnosis



Results

Power and phase-amplitude coupling in PMS

In all frequency bands tested, no differences in relative power were observed between individuals with PMS and TD individuals after correcting for multiple comparisons (FDR=0.1, 5 power comparisons) (Fig. 1). Reduced alpha power in PMS relative to TD approached, but did not reach, significance (U=-4.457, p=0.035). Gamma power enhancements in PMS also did not reach significance (U=2.380, p=0.123).

Mean peak alpha frequency was found to be 9.38 Hz (SD=0.921 Hz) in individuals with PMS, and 9.43 Hz (SD=0.610) in TD individuals; peak alpha frequency was not found to differ between groups (t=0.170, p=0.866). Two individuals with PMS and two TD individuals did not display a peak in alpha activity, and two individuals with PMS demonstrated a peak alpha frequency outside of the 8–12 Hz range (7.32 and 7.81 Hz). These participants all displayed PAC values in the typical range.

When averaging across all channels, individuals with



(4/7) and seizure history (2/7) and there were no significant differences in included vs. excluded participants with PMS on the Vineland Adaptive Behavior Composite, the Vineland Communication Composite, ADOS severity score, NVIQ, SSP, or RBS-R (p > 0.05). All tests were performed using an independent samples Mann–Whitney U test.

PMS largely demonstrated maximal gamma amplitude at the falling phase of the alpha cycle, whereas TD participants largely demonstrated maximal gamma amplitude at the rising phase of the alpha cycle (Fig. 2). Consequently, individuals with PMS demonstrated a positive overall phase bias (median = 4.091×10^{-4} , SD = 1.02×10^{-3}), and TD individuals demonstrated a negative overall phase bias (median = -2.079×10^{-4} , SD = 5.29×10^{-4}) (Fig. 3).



individuals compared to individuals with PMS. Comparisons were done using phase bias values averaged across all channels (Overall), all anterior channels (Anterior), and all posterior channels (Posterior). Median, 10th and 90th percentiles are plotted. *Indicates significance at Benjamini–Hochberg corrected *p* value of .0167

did not demonstrate a difference between groups (U = -0.352, p = 0.738). On the other hand, no differences were found when comparing overall z-MI between groups (Fig. 5, Additional file 1: Table S1). In all participants, no association was found between alpha power and ln(z-MI) (Beta = 0.156, p = 0.337), or phase bias (Beta = -0.240, p = 0.131) when averaging across all electrodes.

Phase-amplitude coupling and clinical characteristics

In all participants, age was significantly associated with $\ln(z-MI)$ (Beta=0.465, p=0.002); age was therefore controlled for in subsequent regressions (Table 2). In a regression comparing $\ln(z-MI)$ and age, the interaction between age and group was not significant (Beta=0.017, p=0.916), indicating the relationship between age and $\ln(z-MI)$ does not differ between individuals with PMS and TD individuals (Fig. 6). In individuals with PMS, $\ln(z-MI)$ increased with RBS-R total score (Beta=0.545, p=0.011); specifically, $\ln(z-MI)$ was found to increase with the Sameness, Ritualistic, and Compulsive sub-



After correcting for multiple comparisons (FDR=0.1, 6 PAC comparisons), overall phase bias was significantly greater in individuals with PMS than TD individuals (U=3.519, p=0.0005).

In both TD individuals and individuals with PMS, phase bias largely increased along the anterior-posterior axis of the scalp, with the exception of electrodes P3 and P4, which demonstrated a negative phase bias in both groups (Fig. 4). Analysis of phase bias in anterior and posterior regions separately showed posterior electrodes exhibited increased phase bias in individuals with PMS (U=2.734, p=0.006) while anterior electrodes did not exhibit a difference between groups (U=0.189, p=0.862) (Figs. 2, 3). Likewise, posterior z-MI was significantly increased in individuals with PMS (U=2.165, p=0.030), while anterior z-MI

scales of the RBS-R (Table 3). No other associations between PAC measures and phenotypic measures reached significance. Additionally, no significant differences in PAC metrics between categorical phenotypic variables within individuals with PMS (sex, ASD diagnosis, *SHANK3* deletion, seizure history) were observed (Table 4).

Discussion

We find individuals with PMS show significantly increased alpha-gamma phase bias relative to TD individuals, with most individuals with PMS demonstrating positive overall phase bias, whereas most typically developing individuals demonstrated negative overall phase bias in our sample. Between-group differences are primarily driven by findings over posterior



Table 2 Associations between behavioral phenotype and PAC metrics in Phelan-McDermid syndrome

Measure	Standardized beta coefficient	<i>P</i> value	
In(z-MI) versus	_	-	
Age	.500	.011*	
Vineland adaptive behavior	.146	.555	
Vineland socialization	040	.870	
Deletion size	092	.723	
SSP total	336	.225	
ADOS comparison score	379	.150	
RBS-R total score	.545	.011*	
NVIQ	.618	.543	
Phase bias versus	-	-	
Age	.194	.342	
Vineland adaptive behavior	104	.630	
Vineland socialization	202	.343	
Deletion size	.232	.261	
SSP total	.367	.100	
ADOS comparison score	.026	.907	
RBS-R total score	092	.652	
NVIQ	980	.338	

All tests were performed as linear regressions, with age included as a control variable for all subsequent tests. A Benjamini–Hochberg correction was applied to In(z-MI) and phase bias comparisons separately. * indicates significance (FDR=.1)

electrodes, where phase bias and PAC are both more strongly positive in individuals with PMS relative to TD individuals. Previous work has reported greater alphagamma PAC in a midline parietal–occipital source in individuals with ASD [24]. Within individuals with



 Table 3
 Associations between RBS-R sub-scales and In(z-MI) in

 Phelan-McDermid Syndrome
 Phelan-McDermid Syndrome

PMS

RBS-R sub-scale	Standardized beta coefficient	<i>P</i> value	
Restricted interest	.337	.183	
Sameness	.546	.012*	
Ritualistic	.490	.019*	
Compulsive	.448	.038*	
Self-injurious	.206	.374	
Stereotypic	.362	.153	

All tests were performed as linear regressions, with age included as a control variable for all subsequent tests. A Benjamini–Hochberg correction was applied. * indicates significance (FDR =.1)

PMS, no differences were observed with measures of overall ASD phenotype, or social functioning; however, RBS-R total score was found to increase with increased PAC strength, indicating in individuals with PMS, PAC strength may map on to this aspect of the ASD symptom profile specifically.

The between-group differences in phase bias suggest that circuit function is perturbed in PMS, in a manner measurable by surface EEG. This finding suggests several opportunities for back-translation into animal models to elucidate underlying mechanisms. For example, scalp level EEG does not reflect the unified activity of the cortex, but rather the grand average of many networks often exhibiting conflicting activity. Phase bias is known to vary by cortical layer. Laminar recordings in monkeys

Measure	PAC median	Mann–Whitney U	P value
z-MI	_	_	_
Sex	_	-	-
Female	2.99 (1.19, 9.83)	2.16	.031
Male	0.836 (0.512, 1.485)	-	-
ASD diagnosis	-	-	-
Yes	1.45 (0.337, 3.73)	712	.501
No	1.60 (0.907, 6.71)	-	-
SHANK3	-	-	-
Deletion	1.22 (0.559, 4.98)	723	.497
Mutation	1.59 (1.23, 9.63)	-	-
Seizure history	-	-	-
Yes	1.84 (.086, 9.50)	.371	.748
No	1.23 (.626, 4.35)	-	-
Phase bias * 10 ⁴	_	-	-
Sex	-	-	-
Female	5.00 (2.29, 15.7)	1.00	.336
Male	3.67 (- 0.007, 5.17)	-	-
ASD diagnosis	_	-	-
Yes	4.28 (2.88, 15.5)	.876	.403
No	3.33 (.534, 7.85)	-	-
SHANK3	_	-	-
Deletion	4.14 (0.210, 15.7)	.145	.910
Mutation	4.04 (0.751, 7.57)	-	-
Seizure history	-	-	_
Yes	7.99 (- 1.85, 18.5)	.889	.409
No	3.84 (1.42, 6.71)	-	-

 Table 4
 Comparing PAC metrics across categorical phenotypes

 in Phelan-McDermid Syndrome
 Phelan-McDermid Syndrome

Median values (25th and 75th percentile values in parentheses) are presented for each category. All tests performed using a Mann–Whitney *U* test. A Benjamini–Hochberg correction was applied to ln(z-MI) and phase bias comparisons separately. * indicates significance (FDR=.1)

and rats demonstrate that spontaneous current sinks in theta and alpha bands in layers 2/3-5a are associated with high gamma amplitudes and high action potential firing (and sources are associated with low gamma amplitudes and low action potential firing) whereas the opposite is true in layer 6 (sinks are associated with low gamma amplitudes and low action potential firing, and sources with high gamma amplitudes and high action potential firing) [31, 55]. Additionally, alpha current generators in layers 2/3 and 6 are in phase with one another, but out of phase with those in layer 4 [55], meaning whether scalp-level EEG gamma activity is phase-locked to the falling phase or the rising phase of alpha could depend on whether alpha activity from layers 2/3 and 6 or layer 4 dominates the signal. Therefore, it is possible the phase bias presented here depends on the relative PAC and alpha activity of each cortical layer. Cortical layer 4 predominantly accepts feedforward (thalamocortical) input, layer 6 predominantly provides feedback (corticothalamic output), and layer 2/3 integrates feedforward, feedback, and lateral activity [56, 57]. Between-group differences in surface level phase bias may therefore suggest altered balance of feedforward versus feedback information transfer in PMS; this could be further examined in animal models.

Here, the phase bias abnormalities in individuals with PMS were localized to electrodes over the posterior cortex. Alpha-gamma PAC has been previously shown to increase in the occipital cortex during visual tasks [58]. Notably, the present study analyzed EEG recordings collected while participants watched a silent movie. Cases of cortical visual impairment have been reported in some individuals with PMS [59]; therefore, the network perturbations captured here may also relate to abnormalities in visual processing in PMS.

Notably, we did not identify any differences in EEG power, in any frequency band, between individuals with PMS and TD individuals. This is in contrast to prior electrophysiological measurements in animal models, where power differences have been demonstrated in specific regions and frequency bands [9, 14, 18]. Still, no consistent findings have emerged in studies of animal models as well as in clinical studies. Here, we used relatively conservative statistical techniques, and a slight relaxing of statistical thresholds would have led to findings of overall low alpha power and high gamma power, consistent with some prior animal studies [9, 14]. Nonetheless, the PAC effects (particularly phase bias) are quite strong and persist despite these conservative techniques. While prior studies of PAC during some tasks demonstrate an inverse relationship between alpha power and PAC [60], we did not identify any such relationship in our sample. This suggests that our PAC findings are not driven by changes in nonsinusoidal alpha activity, and that PAC and alpha power can be independently modulated.

By grossly reflecting neural network activity, EEG is an intermediary on the spectrum from genotype to phenotype. Given the myriad possibilities for analysis that EEG offers, EEG itself thus also reflects a smaller spectrumwithin-a-spectrum from genotype to phenotype, depending on the exact analysis chosen. Here we demonstrate that one analytic technique (phase bias) leans toward a reflection of genotype more than phenotype. Notably, we do not have any data to suggest that this phase bias anomaly is specific to PMS; in fact, it is quite possible that similar phase bias anomalies could be present in other genetic disorders that affect similar pathways (e.g., other disorders of the mTOR pathway), and further research is necessary to test this. On the other hand, our findings suggest that zMI likely measures an aspect of neural network function that leans more towards phenotype; therefore, future research should explore whether zMI anomalies are associated with restricted and repetitive behaviors in other neurodevelopmental disorders. Along similar lines, comparison of PAC and z-MI findings between PMS and a phenotypically similar cohort would also be of interest.

However, the genotype-phenotype spectrum is just one of many axes that EEG-based measures may reflect. For example, EEG-based measures may also change across development. We find that z-MI increases with age in our sample (mean age 9.7 years), but phase bias does not. This extends prior findings, in which z-MI was found to increase across the first 3 years after birth in typical development [33]. Developmental effects are particularly important to consider in PMS given the known molecular and electrophysiological functions of SHANK3, including effects on plasticity. SHANK3 provides scaffolding in the postsynaptic density of glutamatergic synapses [6], and Shank3 mutant mouse models have therefore demonstrated decreased excitability of glutamatergic [8, 9, 14, 15] and GABAergic neurons [11]. Plasticity is also impaired in Shank3 mutants [13]. Excitability can be altered by developmental activity and plasticity within circuits, at times leading to seemingly contradictory findings. For example, when inhibition is impaired more than excitation within corticostriatal circuitry during early development, the balance between activity of excitatory and inhibitory neurons can lead to cortical hyper-activity, with resulting changes in plasticity that ultimately cause high (rather than low) excitability of GABAergic neurons in this circuit [12]. In layer 2/3 of primary somatosensory cortex, Shank3 deficiency causes decreased excitability of GABAergic interneurons but increased excitability of glutamatergic neurons [11]. Trajectories across development, combined with studies examining primary and compensatory mechanisms underlying these trajectories, can provide additional clues about the biological underpinnings of neurodevelopmental disorders including (but not limited to) PMS.

Limitations

Our ability to detect subtle phenotypic associations was hampered by several limitations. First, as is common in rare disease research, the sample size in the PMS group led to limited statistical power for assessing associations with categorical variables within this group. In particular, only 4 PMS individuals exhibited a history of seizures. Additionally, our typically developing cohort was small (15 EEGs analyzed), limiting our ability to identify differences with the PMS group. Second, the severity of PMS led several behavioral measures to suffer from a 'floor' effect, making it difficult to compare the phenotypes of individuals within the PMS group. Clinical assessments were also rarely conducted in in the typically developing cohort; as a result, the present study was not able to test for associations between PAC measures and clinical variables in this group. Also of note, though the individuals with PMS enrolled in this study that were not able to provide adequate EEG data for analysis did not demonstrate clear differences on phenotyping measures, they do represent a subgroup of individuals with PMS this study was not able to capture. Finally, though the scalp-level EEG used here allows us to describe differences in grandaverage oscillatory activity, it is unable to differentiate the specific neural mechanisms underlying these differences; back-translation into animal models will likely be necessary to further explore this.

Conclusion

Altered phase bias in PMS suggests altered network dynamics in this disorder. Mechanisms underlying altered network activity in PMS can be further elucidated using back-translation to determine underpinnings of phase bias abnormalities in animal models.

Future studies may assess the extent to which individuals with other neurodevelopmental and neurogenetic disorders have altered phase bias similar to that seen in PMS, suggesting common underlying mechanisms at the network level, and the extent to which phase bias may change in response to treatment in clinical trials.

Abbreviations

PMS: Phelan-McDermid syndrome; ASD: Autism spectrum disorder; EEG: Electroencephalography; PAC: Phase-amplitude coupling; TD: Typically developing; ADOS: Autism Diagnostic Observation Schedule; SSP: Short sensory profile; RBS-R: Repetitive Behavior Scale-Revised; MSEL: Mullen scales of early learning; SB-5: Stanford Binet-5; DAS-II: Differential ability scales, 2nd edition; NVIQ: Non-verbal intelligence quotient; BEAPP: Batch EEG Automated Processing Platform; HAPPE: Harvard Automated Preprocessing Pipeline for EEG; MI: Modulation index; z-MI: Z-scored Modulation Index; FDR: False discovery rate.

Supplementary Information

The online version contains supplementary material available at https://doi. org/10.1186/s13229-020-00411-9.

Additional file 1. Supplementary Table 1. Group comparisons for power in each frequency band, and phase-amplitude coupling metrics. Median values (25th and 75th percentile values in parentheses) are presented for each category. All tests performed using a Mann-Whitney U test. A Benjamini-Hochberg correction was applied to power and PAC separately. * indicates significance (FDR = .1)

Acknowledgements

The Developmental Synaptopathies Consortium (U54NS092090) is part of the Rare Diseases Clinical Research Network (RDCRN), an initiative of the Office of Rare Diseases Research (ORDR), National Center for Advancing Translational Sciences (NCATS). The content is solely the responsibility of the authors and does not necessarily represent the official views of the National Institutes of Health (NIH). We are grateful to the Boston Children's Hospital IDDRC,

1U54HD090255, for support for this project. We are sincerely indebted to the generosity of the families and patients in PMS clinics across the United States who contributed their time and effort to this study. We would also like to thank the Phelan-McDermid Syndrome Foundation for their continued support in PMS research. Members of the Developmental Synaptopathies Consortium (DSC)—Phelan-McDermid Syndrome Group include:

Mustafa Sahin, MD, PhD ^{*a,b*}, Alexander Kolevzon, MD ^{*c,d*}, Joseph Buxbaum, PhD ^{*c,d,e,f*}, Elizabeth Berry Kravis, MD, PhD ^{*g,h,i*}, Latha Soorya, PhD ^{*j*}, Audrey Thurm, PhD ^{*k*}, Craig Powell, MD, PhD ^{*l,m*}, Jonathan A Bernstein, MD, PhD ^{*n*}, Simon Warfield, PhD ^{*o*}, Benoit Scherrer, PhD ^{*o*}, Rajna Filip-Dhima, MS ^{*a*}, Kira Dies, ScM, CGC ^{*a*}, Paige Siper, PhD ^{*c*}, Ellen Hanson, PhD ^{*p*}, Jennifer M. Phillips, PhD ^{*q*}

Affiliations for above:

a Department of Neurology, Boston Children's Hospital, Harvard Medical School, Boston, MA; b F.M. Kirby Neurobiology Center, Boston Children's Hospital, Harvard Medical School, Boston, MA; c Seaver Autism Center for Research and Treatment, Mount Sinai School of Medicine, New York, NY: d Department of Psychiatry, Mount Sinai School of Medicine, New York, NY; e Department of Genetics and Genomic Sciences, Mount Sinai School of Medicine, New York, NY; f Department of Neuroscience, Mount Sinai School of Medicine, New York, NY; g Department of Pediatrics, Rush University Medical Center, Chicago, IL; h Department of Neurological Sciences, Rush University Medical Center, Chicago, IL; i Department of Biochemistry, Rush University Medical Center, Chicago, IL; j Department of Psychiatry, Rush University Medical Center, Chicago, IL; k Pediatrics and Developmental Neuroscience Branch, National Institute of Mental Health, National Institutes of Health, Bethesda, MD; I Department of Neurology and Neurotherapeutics, University of Texas Southwestern Medical Center, Dallas, TX; m Department of Psychiatry and Neuroscience Graduate Program, University of Texas Southwestern Medical Center, Dallas, TX; n Department of Pediatrics, Stanford University School of Medicine, Stanford, CA o Department of Radiology, Boston Children's Hospital, Harvard Medical School, Boston, MA p Department of Developmental Medicine, Boston Children's Hospital, Harvard Medical School, Boston, MA; q Department of Psychiatry and Behavioral Sciences, Stanford University School of Medicine, Stanford, CA

Authors' contributions

MGM and ARL assisted with study design, data analysis, data interpretation and manuscript preparation. EBK, JDB, LEE, JHFF, AK, MEM, MWM, CMP, PMS, LS, A Thurm and MS contributed to study design and data collection. CAN contributed to study design, data collection, and data interpretation. A Thaliath contributed to data collection. BZ performed statistical analyses. All authors read and approved the final manuscript.

Funding

Research reported in this publication was supported by the National Institute of Neurological Disorders And Stroke of the National Institutes of Health (NINDS), Eunice Kennedy Shriver National Institute Of Child Health and Human Development (NICHD), National Institute Of Mental Health (NIMH) including the Intramural Research Program of the NIMH (1ZICMH002961) and National Center For Advancing Translational Sciences (NCATS), and by the Child Neurology Foundation.

Availability of data and materials

The datasets analyzed during the current study are available from the corresponding author on reasonable request.

Ethics approval and consent to participate

The study was approved by the Institutional Review Board (IRB-P00021244 and IRB-P00013300) at Boston Children's Hospital, which serves as the central IRB for all the sites included in this study. Informed consent was obtained from the legal guardians of all participants and assent was obtained from participants when appropriate.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

Author details

Department of Neurology, Boston Children's Hospital, Boston, MA, USA. ²Department of Pediatrics, Rush University Medical Center, Chicago, IL, USA. ³Department of Neurological Sciences, Rush University Medical Center, Chicago, IL, USA. ⁴Department of Biochemistry, Rush University Medical Center, Chicago, IL, USA. ⁵Seaver Autism Center, Icahn School of Medicine at Mount Sinai Hospital, New York, NY, USA. ⁶Department of Psychiatry, Icahn School of Medicine at Mount Sinai, New York, NY, USA. ⁷Department of Genetics and Genomic Sciences, Mount Sinai School of Medicine, New York, NY, USA. ⁸Department of Neuroscience, Mount Sinai School of Medicine, New York, NY, USA. ⁹Department of Pediatrics, University of Oklahoma Health Science Center, Oklahoma City, OK, USA. ¹⁰Clinical Child Psychology Program, Schiefelbusch Institute for Life Span Studies, University of Kansas, Lawrence, KS, USA. ¹¹Department of Pediatrics, Boston Children's Hospital, Boston, MA, USA. ¹²Department of Neurobiology, UAB School of Medicine, Birmingham, AL, USA. ¹³Department of Psychiatry, Rush University Medical Center, Chicago, IL, USA. ¹⁴Intramural Research Program, National Institute of Mental Health, Bethesda, USA

Received: 22 June 2020 Accepted: 28 December 2020 Published online: 28 April 2021

References

- Nesslinger NJ, Gorski JL, Kurczynski TW, Shapira SK, Siegel-Bartelt J, Dumanski JP, et al. Clinical, cytogenetic, and molecular characterization of seven patients with deletions of chromosome 22q13.3. Am J Hum Genet. 1994;54(3):464–72.
- 2. Phelan MC, Rogers RC, Saul RA, Stapleton GA, Sweet K, McDermid H, et al. 22q13 deletion syndrome. Am J Med Genet. 2001;101(2):91–9.
- Oberman LM, Boccuto L, Cascio L, Sarasua S, Kaufmann WE. Autism spectrum disorder in Phelan-McDermid syndrome: initial characterization and genotype-phenotype correlations. Orphanet J Rare Dis. 2015;10(1):105.
- Soorya L, Kolevzon A, Zweifach J, Lim T, Dobry Y, Schwartz L, et al. Prospective investigation of autism and genotype-phenotype correlations in 22q13 deletion syndrome and SHANK3 deficiency. Mol Autism. 2013;4(1):18.
- Holder JL, Quach MM. The spectrum of epilepsy and electroencephalographic abnormalities due to SHANK3 loss-of-function mutations. Epilepsia. 2016;57(10):1651–9.
- 6. Boeckers TM. The postsynaptic density. Cell Tissue Res. 2006;326:409-22.
- Wang X, Xu Q, Bey AL, Lee Y, Jiang Y. Transcriptional and functional complexity of Shank3 provides a molecular framework to understand the phenotypic heterogeneity of SHANK3 causing autism and Shank3 mutant mice. Mol Autism. 2014;5(1):30.
- Peça J, Feliciano C, Ting JT, Wang W, Wells MF, Venkatraman TN, et al. Shank3 mutant mice display autistic-like behaviours and striatal dysfunction. Nature. 2011;472(7344):437–42.
- Wang X, Bey AL, Katz BM, Badea A, Kim N, David LK, et al. Altered mGluR5-Homer scaffolds and corticostriatal connectivity in a Shank3 complete knockout model of autism. Nat Commun. 2016;7(1):11459.
- Yoo T, Cho H, Park H, Lee J, Kim E. Shank3 Exons 14–16 deletion in glutamatergic neurons leads to social and repetitive behavioral deficits associated with increased cortical layer 2/3 neuronal excitability. Front Cell Neurosci. 2019;10:13.
- Chen Q, Deister CA, Gao X, Guo B, Lynn-Jones T, Chen N, et al. Dysfunction of cortical GABAergic neurons leads to sensory hyper-reactivity in a Shank3 mouse model of ASD. Nat Neurosci. 2020;23(4):520–32.
- Peixoto RT, Wang W, Croney DM, Kozorovitskiy Y, Sabatini BL. Early hyperactivity and precocious maturation of corticostriatal circuits in Shank3B—/— mice. Nat Neurosci. 2016;19(5):716–24.
- Tatavarty V, Torrado Pacheco A, Groves Kuhnle C, Lin H, Koundinya P, Miska NJ, et al. Autism-associated Shank3 is essential for homeostatic compensation in rodent V1. Neuron. 2020;106(5):769–777.e4.
- 14. Yoo Y-E, Yoo T, Lee S, Lee J, Kim D, Han H-M, et al. Shank3 mice carrying the human Q321R mutation display enhanced self-grooming, abnormal electroencephalogram patterns, and suppressed neuronal excitability and seizure susceptibility. Front Mol Neurosci. 2019;18:12.
- Yang M, Bozdagi O, Scattoni ML, Wohr M, Roullet FI, Katz AM, et al. Reduced excitatory neurotransmission and mild autism-relevant

phenotypes in adolescent Shank3 null mutant mice. J Neurosci. 2012;32(19):6525–41.

- Khan OI, Zhou X, Leon J, Kessler R, Gaughan T, D'Souza P, et al. Prospective longitudinal overnight video-EEG evaluation in Phelan–McDermid Syndrome. Epilepsy Behav. 2018;1(80):312–20.
- 17. Spence SJ, Schneider MT. The role of epilepsy and epileptiform EEGs in autism spectrum disorders. Pediatr Res. 2009;65(6):599–606.
- Dhamne SC, Silverman JL, Super CE, Lammers SHT, Hameed MQ, Modi ME, et al. Replicable in vivo physiological and behavioral phenotypes of the Shank3B null mutant mouse model of autism. Mol Autism. 2017;8(1):26.
- Wang J, Barstein J, Ethridge LE, Mosconi MW, Takarae Y, Sweeney JA. Resting state EEG abnormalities in autism spectrum disorders. J Neurodev Disord. 2013;5(1):24.
- Hyafil A, Giraud A-L, Fontolan L, Gutkin B. Neural cross-frequency coupling: connecting architectures, mechanisms, and functions. Trends Neurosci. 2015;38(11):725–40.
- 21. Canolty RT, Knight RT. The functional role of cross-frequency coupling. Trends Cogn Sci. 2010;14(11):506–15.
- 22. Gips B, van der Eerden JPJM, Jensen O. A biologically plausible mechanism for neuronal coding organized by the phase of alpha oscillations. Eur J Neurosci. 2016;44(4):2147–61.
- Port RG, Dipiero MA, Ku M, Liu S, Blaskey L, Kuschner ES, et al. Children with autism spectrum disorder demonstrate regionally specific altered resting-state phase-amplitude coupling. Brain Connect. 2019;9(5):425–36.
- 24. Berman JI, Liu S, Bloy L, Blaskey L, Roberts TPLL, Edgar JC. Alpha-togamma phase-amplitude coupling methods and application to autism spectrum disorder. Brain Connect. 2015;5(2):80–90.
- Khan S, Gramfort A, Shetty NR, Kitzbichler MG, Ganesan S, Moran JM, et al. Local and long-range functional connectivity is reduced in concert in autism spectrum disorders. Proc Natl Acad Sci. 2013;110(8):3107–12.
- Radwan B, Dvorak D, Fenton AA. Impaired cognitive discrimination and discoordination of coupled theta-gamma oscillations in Fmr1 knockout mice. Neurobiol Dis. 2016;1(88):125–38.
- Ethridge LE, White SP, Mosconi MW, Wang J, Pedapati EV, Erickson CA, et al. Neural synchronization deficits linked to cortical hyper-excitability and auditory hypersensitivity in fragile X syndrome. Mol Autism. 2017;8(1):22.
- Robertson CE, Baron-Cohen S. Sensory perception in autism. Nat Rev Neurosci. 2017;18(11):671–84.
- Rosenberg A, Patterson JS, Angelaki DE. A computational perspective on autism. Proc Natl Acad Sci. 2015;112(30):9158–65.
- Pellicano E, Burr D. When the world becomes 'too real': a Bayesian explanation of autistic perception. Trends Cogn Sci. 2012;16(10):504–10.
- Sotero RC, Bortel A, Naaman S, Mocanu VM, Kropf P, Villeneuve M, et al. Laminar distribution of phase-amplitude coupling of spontaneous current sources and sinks. Front Neurosci. 2015;9:454.
- Port RG, Berman JI, Liu S, Featherstone RE, Roberts TPL, Siegel SJ. Parvalbumin cell ablation of NMDA-R1 leads to altered phase, but not amplitude, of gamma-band cross-frequency coupling. Brain Connect. 2019;9(3):263–72.
- Mariscal MG, Levin AR, Gabard-Durnam LJ, Tager-Flusberg H, Nelson CA. Developmental changes in EEG phase amplitude coupling and phase preference over the first three years after birth. bioRxiv. 2019;8(5):818583.
- Soplata AE, McCarthy MM, Sherfey J, Lee S, Purdon PL, Brown EN, et al. Thalamocortical control of propofol phase-amplitude coupling. PLOS Comput Biol. 2017;13(12):e1005879.
- Lega B, Burke J, Jacobs J, Kahana MJ. Slow-theta-to-gamma phase-amplitude coupling in human hippocampus supports the formation of new episodic memories. Cereb Cortex. 2016;26(1):268–78.
- Staudigl T, Hanslmayr S. Theta oscillations at encoding mediate the context-dependent nature of human episodic memory. Curr Biol. 2013;23(12):1101–6.
- Sparrow SS, Balla DA, Cicchetti DV. Vineland-II adaptive behavior scales. Detroit: AGS Publishing; 2005.
- Lord C, Rutter M, DiLavore P, Risi S. Autism diagnostic observation schedule–2nd edition (ADOS-2). Los Angeles: West Psychol Corp; 2012.
- 39. Lord C, Rutter M, Le Couteur A. Autism diagnostic interview-revised: a revised version of a diagnostic interview for caregivers of individuals

with possible pervasive developmental disorders. J Autism Dev Disord. 1994;24(5):659–85.

- American Psychiatric Association. Diagnostic and statistical manual of mental disorders. 5th ed. Washington: American Psychiatric Association; 2013.
- Engel-Yeger B, Habib-Mazawi S, Parush S, Rozenman D, Kessel A, Shani-Adir A. The sensory profile of children with atopic dermatitis as determined by the sensory profile questionnaire. J Am Acad Dermatol. 2007;57(4):610–5.
- Bodfish JW, Symons FJ, Parker DE, Lewis MH. Varieties of repetitive behavior in autism: comparisons to mental retardation. J Autism Dev Disord. 2000;30(3):237–43.
- Em M. Mullen scales of early learning. Circ Pines: MN Am Guid Serv; 1995. p. 207–16.
- Roid G. Stanford-Binet intelligence scales. Itasca: Riverside Publishing; 2003.
- Elliot CD. Differential ability scales-second edition (DAS-II). San Antonio: Harcourt Assessment; 2007.
- Stephens RL, Langworthy B, Short SJ, Goldman BD, Girault JB, Fine JP, et al. Verbal and nonverbal predictors of executive function in early childhood. J Cogn Dev. 2018;19(2):182–200.
- 47. Wang J, Ethridge LE, Mosconi MW, White SP, Binder DK, Pedapati EV, et al. A resting EEG study of neocortical hyperexcitability and altered functional connectivity in fragile X syndrome Refining translational treatment development in fragile X syndrome. J Neurodev Disord. 2017;9(1):11.
- Levin AR, Méndez Leal AS, Gabard-Durnam LJ, O'Leary HM. BEAPP: the batch electroencephalography automated processing platform. Front Neurosci. 2018;7(12):513.
- Gabard-Durnam LJ, Mendez Leal AS, Wilkinson CL, Levin AR. The Harvard automated processing pipeline for electroencephalography (HAPPE): standardized processing software for developmental and high-artifact data. Front Neurosci. 2018;12:1–24.
- Wilkinson CL, Levin AR, Gabard-Durnam LJ, Tager-Flusberg H, Nelson CA. Reduced frontal gamma power at 24 months is associated with better expressive language in toddlers at risk for autism. Autism Res. 2019;12:1211–24.
- 51. Babadi B, Brown EN. A review of multitaper spectral analysis. IEEE Trans Biomed Eng. 2014;61(5):1555–64.
- Tort ABL, Komorowski R, Eichenbaum H, Kopell N. Measuring phaseamplitude coupling between neuronal oscillations of different frequencies. J Neurophysiol. 2010;104(2):1195–210.
- Dupré la Tour T, Tallot L, Grabot L, Doyère V, van Wassenhove V, Grenier Y, et al. Non-linear auto-regressive models for cross-frequency coupling in neural time series. PLOS Comput Biol. 2017;13(12):1005893.
- Canolty RT, Barbaro NM, Edwards E, Kirsch HE, Dalal SS, Nagarajan SS, et al. High gamma power is phase-locked to theta oscillations in human neocortex. Science. 2007;313(5793):1626–8.
- Bollimunta A, Mo J, Schroeder CE, Ding M. Neuronal mechanisms and attentional modulation of corticothalamic alpha oscillations. J Neurosci. 2011;31(13):4935–43.
- Guillery RW, Sherman SM. Thalamic relay functions and their role in corticocortical communication: generalizations from the visual system. Neuron. 2002;33(2):163–75.
- D'Souza RD, Meier AM, Bista P, Wang Q, Burkhalter A. Recruitment of inhibition and excitation across mouse visual cortex depends on the hierarchy of interconnecting areas. Elife. 2016;5:e19332.
- Voytek B. Shifts in gamma phase–amplitude coupling frequency from theta to alpha over posterior cortex during visual tasks. Front Hum Neurosci. 2010;4:191.
- Bosch DGM, Boonstra FN, Reijnders MRF, Pfundt R, Cremers FPM, de Vries BBA. Chromosomal aberrations in cerebral visual impairment. Eur J Paediatr Neurol. 2014;18(6):677–84.
- 60. Jensen O, Spaak E, Park H. Discriminating Valid from Spurious Indices of Phase-Amplitude Coupling. eNeuro. 2016;3(6):ENEURO.0334-16.2016.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.